& ECOHYDROLOGY HYDROBIOLOGY

DOI: 10.2478/v10104-012-0001-7

Vol. 12 No. 1, 35-41 2012

Two different feeding tactics of young-of-the-year perch, *Perca fluviatilis* L., inhabiting the littoral zone of the lowland Sulejow Reservoir (Central Poland)

Piotr Frankiewicz^{1,2}, Adrianna Wojtal-Frankiewicz¹

 ¹ Department of Applied Ecology, University of Lodz, Poland
² International Institute of the Polish Academy of Sciences European Regional Centre for Ecohydrology u/a UNESCO Lodz, Poland

Abstract

The feeding pattern of young-of-the-year (YOY) fish was investigated in a field experiment in the sparsely vegetated littoral zone of the Sulejow Reservoir in June 2007. Perch received special emphasis in this study. During the study period, the part of the reservoir selected to conduct the research was densely inhabited by YOY fish (up to 20 individuals per square metre). The dominant YOY species were perch (*Perca fluviatilis*) and roach (*Rutilus rutilus*). Analyses of stomach/gut contents showed that large zooplankters and benthic prey contributed significantly to the diet of YOY fish. For perch, two distinct feeding tactics were observed. Fish collected by trapping in unvegetated gaps among beds of macrophytes preyed almost exclusively on daphnids and copepods, whereas individuals seined in shallow water among macrophytes ate mostly the larvae of benthic insects. The observed division of YOY perch into two feeding groups may indicate an attempt to decrease the level of both intra-specific competition among the cohort's members and inter-specific competition between perch and roach.

Key words: juvenile fish, littoral, feeding behaviour, zooplankton.

1. Introduction

The foraging behaviour of YOY perch has been the topic of numerous laboratory and field studies (e.g., Persson, Greenberg 1990a; 1990b; Diehl, Eklow 1995; Mehner *et al.* 1998; Hjelm *et al.* 2000; Horppila *et al.* 2000; Wanzenböck *et al.* 2006; Borcherding, Magnhagen 2008). The ontogenetic shift by juveniles from zooplankton to benthic prey is a well-recognised phenomenon. This shift usually serves to optimise the growth rate of YOY fish (Persson, Greenberg 1990b; Byström *et al.* 1998; Hjelm *et al.* 2000). The time of the shift depends not only on the size of the fish but also on the abundance of alternative prey, the strength of inter- and intraspecific competition, and the presence of predators (Persson 1986; Bergman 1990; Diehl 1993; Diehl, Eklow 1995; Sharma, Bystrøm 2008). It has generally been found that YOY perch inhabiting open water eat zooplankton, whereas conspecifics living in the littoral zone feed on benthic prey (Diehl 1992; Hjelm *et al.* 2001; Svanbäck, Eklöv 2008; Kratochvil *et al.* 2008). The division of the YOY perch into two groups differing in feeding pattern even results in changes in the morphological traits of the perch (Hjelm *et al.* 2001; Svanbäck, Eklöv 2002; Borcherding, Magnhagen 2008).

However, investigations of the stomach contents of YOY perch collected from the littoral zone have frequently shown that some individuals do not forage only on benthic prey. Rather, some individuals forage exclusively on zooplankton, and others have mixed diets (Horppila et al. 2000; Okun, Mehner 2005). Possibly, these zooplanktivorous specimens represent a particular fraction of littoral-dwelling fish that are likely to penetrate the water column more actively, forage among macrophytes and specialise in feeding on zooplankton. In contrast, another fraction of the population would consist of benthivorous fish that are less active and spend much more time closer to the bottom. If this hypothesis is true, one would expect that the amount of zooplankton in the stomachs of fish collected by trapping in the water column would be much greater than the amount of zooplankton in the stomachs of fish collected by beach seining. Alternatively, if all YOY perch in the littoral zone exhibit similar activity and frequently change their foraging habitats, the probability of catching fish which stomachs contained zooplankton or benthic prey should be the same no matter where the fish were actually caught.

The aim of the field research presented in this paper was to determine whether food niche diversification actually occurs among YOY perch at a small spatial scale within the littoral zone.

1.1. Study site

The Sulejow Reservoir is a shallow, lowland reservoir situated in central Poland along the middle course of the Pilica River. The maximum length of the reservoir is 15.5 km, and the maximum width is 2.1 km. At maximum capacity $(75 \times 10^6 \text{ m}^3)$, the reservoir covers 22 km², with a mean depth of 3.3 m and a maximum depth of 11 m. The shoreline is approximately 54 km in length. The mean water retention time of the reservoir is 30 days. The Sulejow Reservoir is an eutrophic ecosystem. The mean total phosphorus concentration during the last ten years was approximately 137 µg dm⁻³, ranging from 13 to 1053 µg dm⁻³ (Wagner *et al.* 2009). The dominant species of bloom-forming cyanobacteria is Microcystis aeruginosa Kutzing. These microorganisms produce microcystin-LR, -YR and -RR (Tarczyńska et al. 2001; Jurczak et al. 2005). The mean chlorophyll concentration during growth seasons is approximately 30 mg m⁻³, but it can exceed 100 mg m-3 during phytoplankton blooms (Wagner et al. 2009). The mean summer biomass of zooplankton sampled in the pelagic zone has ranged during last 10 years between 4 mg and 11 mg 1-1 and consists primarily of cladocerans: Daphnia cucullata (Sars), D. longispina (O.F.Müller), Leptodora kindtii (Focke) and Bosmina coregoni (Baird) (Wojtal-Frankiewicz, unpublished data). The adult fish stock is composed mostly of roach (Rutilus rutilus L.), common bream (Abramis brama L.), white bream (Blicca bjoerkna L.), pikeperch (Stizostedion lucioperca L.), perch (Perca fluviatilis L.), bleak (Alburnus alburnus L.) and asp (Aspius aspius L.). Perch and roach dominate the YOY fish community in the littoral zone (Frankiewicz, unpublished data).

Investigations were conducted in a vegetated lacustrine part of the reservoir (Tresta Bay) where the following macrophytes were present: *Potamogeton lucens* L., *Polygonum amphibium* L., *Elodea Canadensis* Michx., *Galium palustre* L., *Carex acuta* L., *Equisetum fluviatile* L., *Eleocharis palustris* (L.) Roem. & Schult., *Gliceria fluitans* (L.) R.Br. and *Iris pseudoacorus* L. The coverage of all species at sample sites was about 50%.

2. Materials and methods

YOY fish were collected on 23-24 June 2007 in the littoral zone using two methods: beach seine netting (to sample all fish from the shallow, vegetated part of the littoral) and plastic traps fixed in the water column (to catch fish active in unvegetated gaps among beds of submerged macrophytes). Five sets of double bottles (of 1.5 dcm³ volume) were placed in water approximately 1 m deep, 30 cm above the bottom. The distance from the shore and between traps was about 10 m and 3 m, respectively. The opening of one bottle was directed towards the bank to trap fish moving away from shore. The opening of the other bottle was directed towards the pelagic zone to trap fish moving inshore (Fig. 1). The bottles were set out at 6 p.m. and then emptied twice, at 11 p.m. and 6 a.m. This time schedule was chosen based on the finding from earlier research in Sulejow Reservoir that intense foraging activity of YOY perch occurred mostly at dusk and dawn (Zalewski et al. 1990). The beach seining was conducted after dusk using 10 m long net with a mesh size of 1 mm.

Fish were anaesthetised and preserved in 10% formalin. Each fish was measured to the nearest 0.5 mm (LT) and weighed to the nearest 0.01 g. The stomach contents of perch were weighed to the nearest 0.1 mg. The prey were counted under a microscope and identified to the lowest practical taxon. The numerical contribution (Hyslop 1980) of a given prey category to the content of each stomach was calculated. The Mann-Whitney U-test was used to compare differences in the number of fish collected in the traps with regard to time (dusk, dawn) and movement direction (inshore, outshore). To test for the habitat effect (traps vs. beach seining) on the fish length, weight, fullness index and the numerical contribution of the main prey categories to

the stomach contents, *t*-test was applied. To test for time and traps orientation effects on the parameters analysed, we performed two-way ANOVA test with the time and traps orientation as categorical factors and individual fish length, weight, fullness index and the numerical contribution of the main prey categories to the stomach contents as dependent variables. Data on prey numbers in perch stomachs were $log_{10}(x + 1)$ transformed to achieve normality and homoscedasticity.

3. Results

The numbers of YOY perch collected in traps were 73 at dusk (41 and 32 in inshore- and outshore-oriented traps, respectively) and 43 at dawn (24 and 19 in inshore- and outshore-oriented traps, respectively). Number of fish in each set of five bottles was highly variable and comparison of trapped fish distribution with regard to both time and movement direction did not reveal significant differences (Mann-Whitney U-test, p = 0.17 and p = 1, respectively). Perch collected by using a beach net had significantly higher length, weight and fullness index than trapped fish (t test: t = 4.03, df = 147, p < 0.001; t = 3.59, df = 147, p < 0.001; t = 3.38, df = 87, p < 0.01, respectively) (Fig. 2). Perch trapped in the morning had significantly lower body weight and stomach fullness than fish trapped at night (two-way ANOVA: $F_{1.112} = 4.33$, p < 0.05; $F_{1.70} = 4.26$, p < 0.05, respectively). Differences in perch length between morning and night samples were not significant (two-way ANOVA: $F_{1,112} = 2.42$, p = 0.12). Perch length, weight and fullness index did not differ significantly between inshore- and outshore-oriented traps (two-way



Fig. 1. Diagram of traps used for collecting YOY perch.

ANOVA: $F_{1,112} = 1.95$, p = 0.17; $F_{1,112} = 0.76$, p = 0.39; $F_{1,70} = 0.03$, p = 0.86, respectively). Time/traps orientation interactions were not significant (two-way ANOVA: $F_{1,112} = 1.95$, p = 0.17; $F_{1,112} = 0.76$, p = 0.39; $F_{1,70} = 0.03$, p = 0.86, for perch length, weight and fullness index, respectively).

The numerical contribution of the main prey categories to the stomach content of YOY perch differed strongly between trapped fish and fish collected by using a beach net (Fig. 3). Daphnids and predatory cladocerans, *Leptodora kindtii*, were not present in the stomachs of seined perch. The numbers of copepods found in perch stomachs obtained from trap samples were significantly higher than those found in the seine sample (*t* test: t = 2.87, df = 81, p < 0.01) (Fig. 3). Unlike trapped perch, seined fish preyed mainly upon benthic insects, mostly chironomid larvae (Fig. 3). These differences were highly significant (*t* test: t = 8.56, df = 81, p < 0.001).

L. kindtii were significantly more numerous in the stomachs of perch trapped at night than in the stomachs of fish trapped in the morning (two-way ANOVA: F_{1.67} = 82.15; p < 0.001) (Fig. 3). Significant differences were also found for Daphnia sp. These filtering zooplankters were much more numerous in the food of perch entering traps from the open water than in the food of perch entering traps from the bank (two-way ANOVA: $F_{1.67} = 10.44$; p < 0.01) (Fig. 3). Copepods were the other noteworthy group of zooplankton whose contribution to the food of the perch varied significantly. They were more numerous in the stomachs of perch trapped at night than in the stomachs of fish trapped in the morning (two-way ANOVA: $F_{1,67} = 19.01$; p < 0.001) (Fig. 3). Time/traps orientation interactions were not significant for any prey categories. Differences in the contribution of benthic organisms to the food of trapped perch were not analyzed due to the low number of these prey.

4. Discussion

The feeding ecology of YOY perch has already been well characterised in different freshwater ecosystems (e.g. Persson, Greenberg 1990a; 1990b; Diehl, Eklow 1995; Mehner *et al.* 1998; Hjelm *et al.* 2000; Horppila *et al.* 2000; Vašek *et al.* 2006; Borcherding, Magnhagen 2008). The most characteristic life-history phenomena described for these fish are the presence of horizontal migration from the littoral to the pelagic zone and back during the first months of life (Wang, Eckman 1994; Urho 1996) and the presence of ontogenetic shifts in the utilisation of food resources, from zooplankton to benthos and finally to fish (Thorpe 1977; Persson 1990; Hjelm *et al.* 2000). The occurrence and timing of this ontogenetic shift are highly variable and



Fig. 2. Comparison of length, weight and fullness index of YOY perch from traps and from beach seining (boxes indicate means values, whiskers indicate standard deviations).

(+) – traps directed towards the bank; (-) – traps directed towards the open water.



Fig. 3. Contribution (by number) of the main food categories of YOY perch from traps and from beach seining (boxes indicate means values, whiskers indicate standard errors).

APN – average prey number by fish; N_{fish} – number of fish used for food analyses. (+) – traps directed towards the bank; (-) – traps directed towards the open water.

depend mostly on prey availability (Persson 1986; Byström et al. 1998), on the intensity of inter- and intraspecific competition (Bergman 1990; Persson, Greenberg 1990a, 1990b; Diehl 1993), and on the threat of predation (Persson 1991; Diehl, Eklow 1995; Sharma, Bystrøm 2008). The need to adjust to a complex pressure of environmental stressors results in the highly diverse feeding pattern of YOY perch reported in numerous studies (Zalewski et al. 1990; 2006; Kahl, Radke 2006; Quevedo, Olsson 2006; Olsson et al. 2007; Huss et al. 2008; Urbatzka et al. 2008). However, some general patterns may be described. The main food of YOY perch foraging in the pelagic zone consists of zooplankton and is dominated by daphnids (Svanbäck, Eklow 2002; Vašek et al. 2006). Benthic macroinvertebrates are usually predominant in the stomach contents of perch foraging in the littoral zone (Zalewski et al. 1990; Okun, Mehner 2005). In the littoral zone, the relative contribution of benthic prey to the diet

of YOY perch should increase with the size of the fish (Guma'a 1978; Treasurer 1990). However, in the presence of competitors (e.g. YOY roach) they may be forced to use benthic prey at a fairly small size (Persson, Greenberg 1990a, 1990b). The time of the known shift to benthivory also depends on the relative abundance of benthic macroinvertebrates and zooplankton (Persson, Greenberg 1990b) as well as on the presence of superior competitors for benthic prey, like ruffe and/or older conspecifics (Bergman, Greenberg 1994). Generally, the growth rate of juvenile perch is higher if large cladocerans are available (e.g., Mehner et al. 1995; Romare 2000). Thus, an early shift towards benthic food may reflect unfavourable conditions for perch (Persson 1986; Byström et al. 1998). The finding that YOY perch released from competition with roach continued to feed on zooplankton through the autumn may support this hypothesis (Persson 1986). However, our observation that YOY perch of a size typical for the zooplanktivorous stage and feeding on zooplankton were smaller than those eating benthic prey demonstrates the unpredictability of foraging behaviour. Actual foraging patterns seem to depend on subtle differences in prey availability and in the overall biotic background.

This research and our previous studies (Wojtal *et al.* 2003) have demonstrated that different fractions of the YOY perch specialise on different food resources on a small spatial scale. This finding is in agreement with the conclusions of Quevedo, Olsson (2006) that large variations in the isotope composition of perch individuals caught in the littoral zone of a mesotrophic lake must result from enduring utilisation of different prey categories at the microhabitat scale.

It should be emphasized that our conclusions about differences between the stomach contents of trapped and seined perch are rather conservative. It is not unlikely that some YOY perch from the "zooplanktivorous fraction" were also caught during beach seining.

Perch foraging in littoral areas face competition with two species of superior competitors. Juvenile roach are superior competitors for zooplankton (e.g. Persson 1987), and ruffe are superior competitors for benthic prey (Bergman, Greenberg 1994; Schleuter, Eckmann 2006). In years of high juvenile perch density, intraspecific competition may also have strong influence on the feeding behaviour of the perch (Persson, Greenberg 1990a; Svanbäck, Persson 2004). Such circumstances may lead to many possible responses that can serve to diminish inter- and intraspecific competition. One such response is the spatial segregation of juvenile perch between littoral and pelagic zone, a phenomenon that has been frequently reported (e.g. Post et al. 1997; Hjelm et al. 2001).

Another possible response to such competition is the performance of diurnal migrations between these two zones (e.g. Gliwicz, Jachner 1992). Because the numbers of fish entering traps from the open water and from the bank did not differ, it is unlikely that the differences in stomach contents observed in our studies resulted from the diurnal migration of YOY perch. Moreover, assuming that zooplanktivorous perch were feeding in the pelagic zone during the day and stayed in the littoral during the night, they should not be trapped with recently eaten (undigested) zooplankton in the morning in the littoral zone. By analogy, if these fish stayed in the pelagic zone during the night, fed there at dusk and dawn and hid in the littoral during the day, they would not be caught at night with their stomachs full of zooplankton. Therefore, trapped fish more likely represented the fraction of perch permanently foraging on zooplankton above the bottom and entering traps by chance.

Overall, our data show that individual-based perch foraging decisions do in fact result in food niche diversification at a low spatial scale inside the littoral zone, at least at high YOY densities.

Acknowledgements

This publication is based upon work conducted at the Sulejowski Reservoir LTER site.

References

- Bergman, E. 1990. Effect of roach, *Rutilus rutilus*, on two percids, *Perca fluviatilis* and *Gymnocephalus cernua*: importance of species interactions for diet schifts. *Oikos* 57, 241-249.
- Bergman, E., Greenberg, L. 1994. Competition between a planktivore, a benthivore, and a species with ontogenetic diet shifts. *Ecology* **75**, 1233-1245.
- Borcherding, J. 2006. Prey or predator: 0+ perch (*Perca fluviatilis*) in the trade-off between food and shelter. *Environmental Biology of Fish* 77, 87-96.
- Borcherding, J., Magnhagen, C. 2008. Food abundance affects both morphology and behaviour of juvenile perch. *Ecology of Freshwater Fish* **17**, 207-218.
- Byström, P., Persson, L., Wahlström, E. 1998. Competing predators and prey: juvenile bottlenecks in whole-lake experiments. *Ecology* 79, 2153-2167.
- Diehl, S. 1992. Fish predation and benthic community structure: the role of omnivory and habitat complexity. *Ecology* **73**, 1646-1661.
- Diehl, S. 1995. Effects of habitat structure on resource availability, diet, and growth of benthivorous perch, *Perca fluviatilis. Oikos* 67, 403-414.
- Diehl, S., Eklöv, P. 1995. Effects of piscivore-mediated habitat use on resources, diet, and growth of perch. *Ecology* 76, 1712-1726.
- Gliwicz, Z.M., Jachner, A. 1992. Diel migrations of juvenile fish: a ghost of predation past or present? *Archive für Hydrobiology* **124**, 385-410.
- Guma'a, S.A. 1978. The food and feeding habits of young perch, *Perca fluviatilis*, in Windermere. *Freshwater Biology* 8, 177-187.
- Hjelm, J., Persson, L., Christensen, B. 2000. Growth, morphological variation and ontogenetic niche shifts in perch (*Perca fluviatilis*) in relation to resource availability. *Oecologia* **122**, 190-199.
- Hjelm, J., Svanbäck, R., Byström, P., Persson, L., Wahlström, E. 2001. Diet-dependent body morphology and ontogenetic reaction norms in Eurasian perch. *Oikos* 95, 311-323.
- Horppila, J., Ruuhijärvi, J., Rask, M., Karppinen, C., Nyberg, K., Olin, M. 2000. Seasonal changes in the diets and relative abundances of perch and roach in the littoral and pelagic zones of a large lake. *Journal* of Fish Biology 56, 51-72.
- Huss, M., Byström, P., Persson, L. 2008. Resource heterogeneity, diet shifts and intra-cohort competition: effects on size divergence in YOY fish. *Oecologia* 158, 249-257.
- Hyslop, E.J. 1980. Stomach contents analysis a review of methods and their application. *Journal of Fish Biology* **17**, 411-429.

- Jurczak, T., Tarczyńska, M., Izydorczyk, K., Mankiewicz, J., Zalewski, M., Meriluoto, J. 2005. Elimination of microcystins by water treatment process – examples from Sulejow Reservoir, Poland. *Water Research* 39, 2394-2406.
- Kahl, U., Radke, R.J. 2006. Habitat and food resource use of perch and roach in a deep mesotrophic reservoir: enough space to avoid competition? *Ecology of Freshwater Fish* 15, 48-56.
- Kratochvil, M., Peterka, J., Kubečka, J., Matěna, J., Vašek, M., Vaničková, I., Čech, M., Sed'a, J. 2008. Diet of larvae and juvenile perch, *Perca fluviatilis* performing diel vertical migrations in a deep reservoir. *Folia Zoologica* 57, 313-323.
- Mehner, T., Schultz, H., Herbst, R. 1995. Interaction of zooplankton dynamics and diet of 0+ perch (*Perca fluviatilis* L.) in the top-down manipulated Bautzen reservoir (Saxony, Germany) during summer. *Limnologica* 25, 1-9.
- Mehner, T., Plewa, M., Hülsmannn, S., Worischka, S. 1998. Gape-size dependent feeding of age-0 perch (*Perca fluviatilis*) and age-0 zander (*Stizostedion lucioperca*) on *Daphnia galeata*. Archiv für Hydrobiologie 142, 191-207.
- Okun, N., Mehner, T. 2005. Distribution and feeding of juvenile fish on invertebrates in littoral reed (*Phragmites*) stands. *Ecology of Freshwater Fish* 14, 139-149.
- Olsson, J., Svanbäck, R., Eklöv, P. 2007. Efects of resource level and habitat type on behavioural and morphological plasticity in Eurasian perch. *Oecologia* 152, 48-56.
- Persson, L. 1986. Effects of reduced interspecific competition on resource utilization in perch (*Perca fluviatilis*). Ecology 67, 355-364.
- Persson, L. 1987. Effects of habitat and season on competitive interactions between roach (*Rutilus rutilus*) and perch (*Perca fluviatilis*). *Oecologia* 73, 170-177.
- Persson, L. 1990. Predicting ontogenetic niche shifts in the field: what can be gained from foraging theory? In: Hughes, R.N. [Ed.] *Behavioural Mechanisms of Food Selection*. Springer, New York, p. 203-218.
- Persson, L. 1991. Behavioral response to predators reverses the outcome of competition between prey species. *Behavioral Ecology and Sociobiology* 28, 101-105.
- Persson, L., Greenberg, L. 1990a. Juvenile competitive bottlenecks: the perch (*Perca fluviatilis*) – roach (*Rutilus rutilus*) interaction. *Ecology* **71**, 44-56.
- Persson, L., Greenberg, L. 1990b. Interspecific and intraspecific size class competition affecting resource use and growth of perch, *Perca fluviatilis*. *Oikos* 59, 97-106.
- Post, J.R., Johannes, M.R.S., McQueen, D.J. 1997. Evidence of density-dependent cohort splitting in age-0 yellow perch (*Perca flavescens*): potential behavioural mechanisms and population-level consequences. *Canadian Journal of Fisheries and Aquatic Sciences* 54, 867-875.
- Romare, P. 2000. Growth of larval and juvenile *perch*: the importance of diet and fish density. *Journal of Fish Biology* 56, 876-889.

- Quevedo, M., Olsson, J. 2006. The effect of small-scale resource origin on trophic position estimates in *Perca fluviatilis*. *Journal of Fish Biology* **69**, 141-150.
- Sharma, C.M., Borgstrøm, R. 2008. Shift in density, habitat use, and diet of perch and roach: an effect of changed predation pressure after manipulation of pike. *Fisheries Research* **91**, 98-106.
- Schleuter, D., Eckmann, R. 2006. Competition between perch and ruffe: the advantage of turning night into day. *Freshwater Biology* **51**, 287-297.
- Svanbäck, R., Eklöv, P. 2002. Effects of habitat and food resources on morphology and ontogenetic trajectories in perch. *Oecologia* 131, 61-70.
- Svanbäck, R., Persson, L. 2004. Individual specialization, niche width and population dynamics: implications for trophic polymorphisms. *Journal of Animal Ecol*ogy **73**, 973-982.
- Tarczyńska, M., Romanowska-Duda, Z., Jurczak, T., Zalewski, M. 2001. Toxic cyanobacterial blooms in drinking water reservoir – causes, consequences and management strategy. *Water Science and Technology Water Supply* 1, 237-246.
- Thorpe, J.E. 1977. Morphology, physiology, behavior and ecology of *Perca fluviatilis* L. and *Percu flavescens* Mitchill. *Journal of Fishery Research Board Canada* 34, 1504-1514.
- Treasurer, J.W. 1990. The food and daily food consumption of lacustrine 0+ perch, *Perca fluviatilis* L. *Freshwater Biology* 24, 361-374.
- Urbatzka, R, Beeck, P., van der Velde, G., Borcherding, J. 2008. Alternative use of food resources causes intra-cohort variation in the size distribution of young-of-the-year perch (*Perca fluviatilis*). Ecology of Freshwater Fish **17**, 475-480.
- Urho, L. 1996. Habitat shifts of perch larvae as survival strategy. Annales Zoologici Fennici 33, 329-340.
- Vašek, M., Kubečka, J., Matěna, J., Sed'a, J. 2006. Distribution and diet of 0+ fish within a canyon-shaped European reservoir in late summer. *International Review of Hydrobiology* **91**, 178-194.
- Wagner, I., Izydorczyk, K., Kiedrzyńska, E., Mankiewicz-Boczek, J., Jurczak, T., Bednarek, A., Wojtal-Frankiewicz, A., Frankiewicz, P., Ratajski, S., Kaczkowski, Z., Zalewski, M. 2009. Ecohydrological system solution for enhancement of ecosystem services: the Pilica river demonstration project. *Ecohydrology & Hydrobiology* 9, 13-39.
- Wang, N., Eckmann, R. 1994. Distribution of perch (*Perca fluviatilis* L.) during their first year of life in Lake Constance. *Hydrobiologia* 277, 135-143.
- Wanzenböck, J., Mikheev, V.N., Pasternak, A.F. 2006. Modification of 0+ perch foraging behaviour by indirect cues of predation risk. *Ecology of Freshwater Fish* 15, 118-124.
- Wojtal, A., Frankiewicz, P., Izydorczyk, K., Zalewski, M. 2003. Horizontal migration of zooplankton in a littoral zone of the lowland Sulejow Reservoir (Central Poland). *Hydrobiologia* **506/509**, 339-346.
- Zalewski, M., Brewińska-Zaraś, B., Frankiewicz P., Kalinowski, S. 1990. The potential for biomanipulation using fry communities in a lowland reservoir: concordance between water quality and optimal recruitment. *Hydrobiologia* 200/201, 549-556.